UNAMBIGUOUS SYNTHESIS OF $(1 \rightarrow 2)$ - AND $(1 \rightarrow 3)$ - RHAMNOPYRANOSYL-RHAMNO-PYRANOSE DERIVATIVES AND THEIR ¹³C-NMR STUDY

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Rhamnobioses are widely distributed in nature and are commonly found as structural units in glycolipids^{1,2} and bacterial polysaccharides^{3,4} and as constituents of glycosides⁵. In order to study the immunological properties of these oligosaccharides and for synthesizing naturally occurring flavonol-glycosides, we have prepared a series of oligosaccharides containing L-rhamnose⁶⁻⁹, including all three 0- α -L-rhamnopyranosyl-L-rhamnoses. Up to now from the rhamnobioses only two unambiguous syntheses^{6,10} of the 4- $0-\alpha$ -L-rhamnopyranosyl-L

Recently, also, the synthesis of the 2-0- α -L-rhamnopyranosyl-L-rhamnopyranose¹¹ and 3-0- α -L-rhamnopyranosyl-L-rhamnopyranose¹² has been reported by different authors. They used modified versions of the "open chain oligosac-charide synthesis", introduced by King and Bishop¹³. Since this method al-ways results in a mixture of different structural isomers, which have to be separated by column chromatography, the above two syntheses cannot be accepted as a proof of structure. Furthermore it is necessary to point out that the main routes of the two syntheses were opposite; i.e. in the case of benzyl 4-0-benzyl- β -L-rhamnopyranoside the main product of the reaction with α -acetobromo-L-rhamnose was the 2-0- α -L-rhamnopyranosyl-L-rhamnose¹¹ deriva- $\frac{741}{741}$

Assignment of the 13 C chemical shifts and $^{1}J_{
m CH}$ data

| Carbon | Compound | | | | | | | | | |
|--|----------|------|-------|----------|----------|----------|-------|-------|-----------|------------|
| | 1 | 2 | 2 | <u>4</u> | <u>6</u> | <u>7</u> | 8 | 2 | <u>10</u> | <u>11</u> |
| 1 | 92.8 | 98.0 | 100.5 | 98.3 | 100.0 | 98.5 | 99.8 | 98.7 | 100.5 | 101.6 |
| ¹ _{J_{CH}} | | | | | | (167) | (164) | (171) | (170) | (170.2) |
| 2 | 75.7 | 78.3 | 68.6 | 78.9 | 76.0 | 78.1 | 76.9 | 71.2 | 79.0 | 70.8 |
| 3 | 79.7 | 78.3 | 80.2 | 71.7 | 79.9 | 78.5 | 70.8 | 75.1 | 70.9 | 78.8 |
| 4 | 78.0 | 81.2 | 80.2 | 82.2 | 80.3 | 80.9 | 71.5 | 72.8 | 73.1 | 72.2 |
| 5 | 64.2 | 64.3 | 67.4 | 67.3 | 68.2 | 68.3 | 67.2 | 64.7 | 69.2 | 69.4 |
| 6 | 17.9 | 17.8 | 18.0 | 18.1 | 18.0 | 18.1 | 17.6 | 17.5 | 17.7 | 17.8 |
| 1' | | | | | 99.3 | 99.4 | 99.4 | 99.0 | 102.9 | 102.9 |
| ¹ J CH | | | | | | (174) | (172) | (173) | (170. | 6) (170.3) |
| 2, | | | | | 69.3 | 69.3 | 68.9 | 68.8 | 71.1 | 71.0 |
| 3' | | | | | 69.9 | 70.1 | 70.2 | 70.6 | 70.9 | 71.1 |
| 4, | | | | | 71.3 | 71.3 | 71.3 | 71.4 | 72.9 | 73.0 |
| 5' | | | | | 66.9 | 67.0 | 66.5 | 66.7 | 69.7 | 69.6 |
| 6, | | | | | 17.4 | 17.5 | 17.3 | 17.3 | 17.8 | 17.8 |

 $^{1}J_{CH}$ coupling constants are given in brakets (Hz). All chemical shifts are referred to TMS (i). For D₂O solutions the dioxane (i) line was used (67.3 ppm).

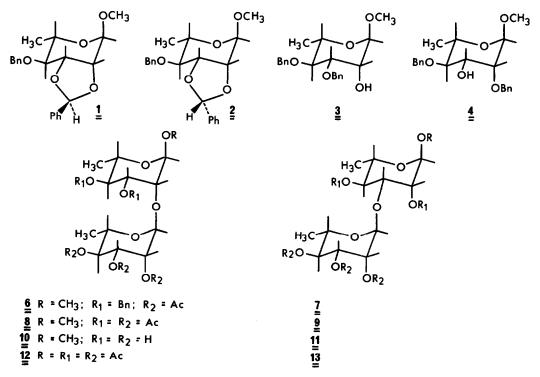


Table.

tive, while the methyl α -L-rhamnopyranoside reacted with α -acetobromo-L-rhamnose to a 3-0- α -L-rhamnopyranosyl-L-rhamnose¹² derivative.

Here we describe the unambiguous synthesis of 2-0- and 3-0- α -L-rhamopyranosyl-L-rhamnopyranose starting from methyl 3,4-di-O-benzyl- α -L-rhamnopyranoside (3) and methyl 2,4-di-0-benzyl- α -L-rhamnopyranoside (4)respectively. <u>3</u> was prepared by the hydrogenolysis of methyl 4-0-benzyl-<u>exo</u>-2,3-0-benzylidene- α -L-rhamnopyranoside $(\underline{1})^{14}$ with LiAlH₄-AlCl₃ reagent¹⁵ with the following data ($\underline{3}$, 98%, syrup, $\left[\alpha\right]_{D}$ -46.4° (CHCl₃). The hydrogenolysis of methyl 4-0-benzyl-<u>endo</u>-2,3-0-benzylidene- α -L-rhamnopyranoside ($\underline{2}$)¹⁴ under the same reaction conditions gave $\frac{4}{2}$ (96%, syrup, $\left[\alpha\right]_{D}$ -15.4° (CHCl₃). Compound <u>3</u> and $\underline{4}$ were coupled with α -acetobromo-L-rhamnose (5) in benzene-nitromethane (1:1) as solvent and in the presence of $Hg(CN)_2$ to obtain after purification $\frac{6}{2}$ (62.5%, m.p. 96-98°C, $\left[\alpha\right]_{D}$ -38.4° (CHCl₃) and $\frac{7}{2}$ (78.8%, syrup, $\left[\alpha\right]_{D}$ -46.1° (CHCl₃), respectively. Catalytic debenzylation of $\underline{6}$ and $\underline{7}$ over Pd/C and subsequent acetylation resulted in methyl 3,4-di-0-acetyl-2-0-(2,3,4-tri-0acetyl-a-L-rhamnopyranosyl)-a-L-rhamnopyranoside (8) (92.7%, m.p. 149-150°C, $\left[\alpha\right]_{D}$ -44.8° (CHCl₃), and methyl 2,4-di-0-acetyl-3-0-(2,3,4-tri-0-acetyl- α -Lrhamnopyranosyl)- α -L-rhamnopyranoside ($\underline{9}$) (95%, m.p. 135-136°C, $\left[\alpha\right]_{D}$ -43.6° (CHCl₃), lit.¹² m.p. 125°C, $\left[\alpha\right]_{D}$ -48° (CHCl₃), respectively. Deacetylation of $\underline{8}$ and $\underline{9}$ gave methyl 2-0- α -L-rhamnopyranosyl- α -L-rhamnopyranoside ($\underline{10}$, 96%, syrup, $\left[\alpha\right]_{D}$ -92° (H₂0), and methyl 3-0- α -L-rhamnopyranosyl- α -L-rhamnopyranoside (11, 95%, syrup, $\left[\alpha\right]_{D}$ -78° (H₂0), respectively. Acetolysis of compounds 8 and 9 yielded the hexa-O-acetyl-dirhamnoses (12 and 13) as anomeric mixtures at the reducing rhamnopyranose ring. The predominating anomers were identified as α -anomers ($\underline{12}$: 62%, m.p. 118-120°C, $\left[\alpha\right]_{D}$ -48° $(CHC1_3); \underline{13}: 65\%, \text{m.p. } 75-76^{\circ}C, [\alpha]_D -38^{\circ} (CHC1_3), \text{ lit.}^{12} \text{ syrup, } [\alpha]_D -32^{\circ}$ (CHCl₃). The structures of compounds $\frac{1}{2} - \frac{4}{2}$, and $\frac{6}{2} - \frac{1}{2}$ were verified by ¹Hand ¹³C-NMR spectroscopy. The ${}^{1}J_{CH}$ coupling constants were measured for <u>8</u> -11 to determine the anomeric configuration of the interglycosidic bonds. They were all found to be α -L (1C), (see Table).

The hydrogenolysis of dioxane- and dioxolane-type benzylidene derivatives opens a general method for preparing selectively blocked sugar derivatives as starting material for the synthesis of complex oligosaccharides. Further investigations are in progress.

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REFERENCES

- 1) S.N. Khanna and P.C. Gupta, Phytochemistry, 6 (1967) 735.
- 2) F.G. Jarvis and M.J. Johnson, <u>J. Amer. Chem. Soc.</u>, 71 (1949) 4124.
- B. Lindberg, J. Lönngren, J.L. Thompson, and W. Nimmich, <u>Carbohydr. Res.</u>, 25 (1972) 49.
- 4) Y.N. Choy and G.G.S. Dutton, Can. J. Chem., 52 (1974) 684.
- 5) H. Wagner, M. Ertan, and O. Seligmann, Phytochemistry, 13 (1974) 857.
- A. Lipták, V.M. Chari, B. Kreil, and H. Wagner, <u>Phytochemistry</u>, 17 (1978) 977.
- 7) A. Lipták and P. Nánási, Carbohydr. Res., 44 (1975) 313.
- 8) H. Wagner, A. Lipták, and P. Nánási, Acta Chim. Hung., 89 (1976) 405.
- 9) A. Lipták and P. Nánási, Tetrahedron Lett., (1977) 921.
- G.M. Bebault, G.G.S. Dutton, and C.K. Warfield, <u>Carbohydr. Res.</u>, 34 (1974) 174.
- W. Schalch, W. Hochstrasser, and D.G. Braun, Tetrahedron Lett., (1978) 4153.
- 12) C. Laffite, A.M.N.P. Du, F. Winternitz, R. Wylde, and F. Pratviel-Sosa, <u>Carbohydr. Res.</u>, 67 (1978) 91.
- R.R. King and C.T. Bishop, <u>Carbohydr. Res.</u>, 32 (1974) 239; <u>Can. J. Chem.</u>, 52 (1974) 3913.
- 14) D.M. Clode, D. Horton, and W. Weckerle, Carbohydr. Res., 49 (1976) 305.
- 15) A. Lipták, <u>Tetrahedron Lett.</u>, (1976) 3551; <u>Carbohydr. Res.</u>, 63 (1978) 69.
- 16) Values of $[\alpha]_D$ were measured with a Perkin-Elmer 241 automatic polarimeter. ¹H-NMR spectra were measured with a Jeol MH-100 spectrometer, and ¹³C-NMR spectra with a Varian XL-100-FT-15 spectrometer in 10-25% (w/v) CDCl₃ or D₂O solutions at 50°C. ¹J_{CH} coupling constants were determined from natural-abundance proton coupled spectra by gated decoupling technique.

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